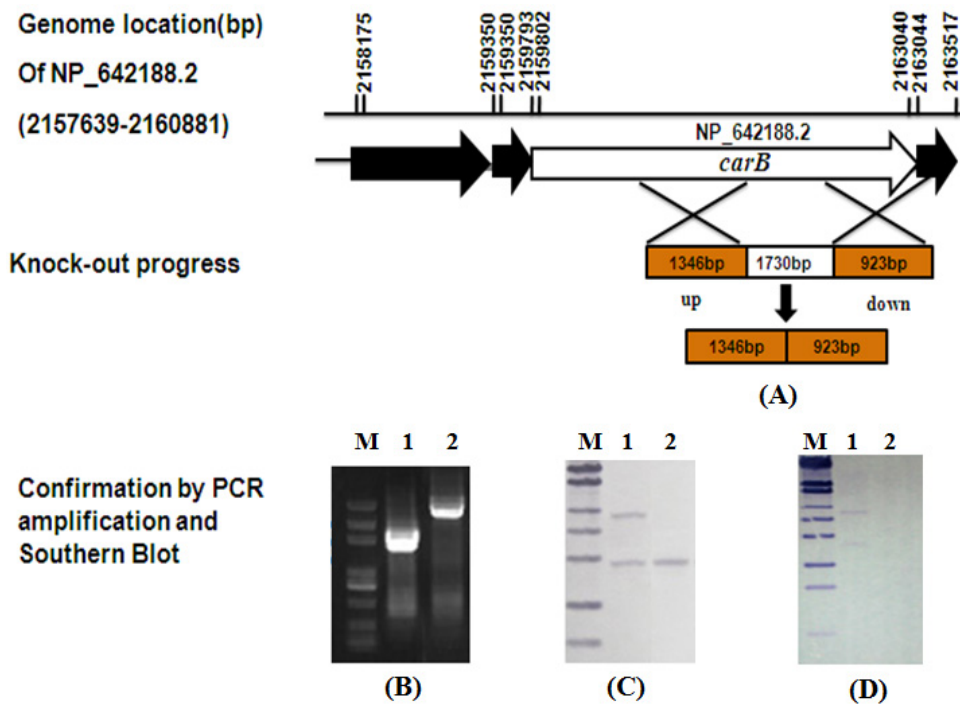


Appendix A Schematic map and molecular analysis of *carA* mutant of *Xanthomonas citri* subsp. *citri*. (A) Schematic process for construction of *carA* knock out mutant from *Xac* 29-1. The 420 bp left flanking fragment and 594 bp right flanking fragment were cloned into pK18mobSacB vector. The *carA* gene was knocked out through two-steps of homology recombination. (B) PCR analysis of *carA* mutant. Lane M, DL5000 marker; Lane 1, Wild type *Xac* 29-1; Lane 2, the *carA* mutant $\Delta carA$. (C) Southern blot for *carA* mutant. The Southern blot was carried out by using the 420 bp upside fragment after the genomic DNA was digested by *Pst* I. Lane M, λ -EcoT14 I marker; Lane 1, Wild type *Xac* 29-1; Lane 2, the *carA* mutant $\Delta carA$.



Appendix B Schematic map and molecular analysis of *carA* and *carAB* mutant of *Xanthomonas citri* subsp. *citri*. (A) Schematic process for construction of *carB* knock out mutant from *Xac* 29-1. The 1346 bp left flanking fragment and 923 bp right flanking fragment were cloned into pK18mobSacB vector. The *carB* gene was knocked out through two-steps of homology recombination. (B) PCR analysis of *carB* mutant. Lane M, DL5000 marker; Lane 1, the *carB* mutant $\Delta carB$; Lane 2, Wild type *Xac* 29-1. (C) Southern blot for *carB* mutant. The Southern blot was carried out by using left flanking fragment as probe after the genomic DNA was digested by *Pst* I. Lane M, λ -EcoT14 I marker; Lane 1, Wild type *Xac* 29-1; Lane 2, the *carB* mutant $\Delta carB$. (D) Southern blot for *carA* and *carB* double mutant. The recombinant construct for *carA* mutagenesis was introduced into $\Delta carB$ genetic background to produce a double mutant. The Southern blot was carried out by using the 723 bp DNA fragment of *carA* gene as probe after the genomic DNA was digested by *Pst* I. Lane M, λ -EcoT14 I marker; Lane 1, Wild type *Xac* 29-1; Lane 2, the double mutant $\Delta carAB$.

Appendix C Primers used for molecular cloning in this study

Primer	Sequence (5'-3')	Description
carA1.F	GCCTCTAGATCGAATCGCATCACGTGCACAA	A 420 bp fragment left to <i>carA</i>
carA1.R	GGTTCGGGCAAGCGAGAATTGTAAA	
carA2.F	TTTACAATTCTCGCTTGCCCGAACCTCGCTGTTTCGATGGCACCAA	A 594 bp fragment right to <i>carA</i>
carA2.R	AATCTGCAGGTCCTCAGTAAAGTCGGTCC	
carB1.F	GGCTCTAGATCAACTGGCAGACGGTCGAAAAG	A 1346 bp fragment left to <i>carB</i>
carB1.R	CAGCGTGCGTACCGATTCTT	
carB2.F	AAGAATCGGTACGCACGCTGGAAGAGCTGGAACACCTGAAGTC	A 923 bp fragment right to <i>carB</i>
carB2.R	AAAACCTGCAGGGTGCTCGAACCAAGTCGTCGCCAAT	
ScarA.F	CCATGACCGGCTATCAGGAAGT	723 bp from <i>carA</i> as a probe for Southern blot
ScarA.R	CTTCTGCGGATCAGTTCCTTG	
carA.F	TATAAGCTTACGTCGCGGCCTATTCATGCCTCAA	A 1409 bp <i>carA</i> gene
carA.R	TTTTCTAGACGGCAGCGAGGCCAGCCCTGTTTTTC	
carB.F	TATTCTAGATGCCCCGATCGCCCGACCAAATTACC	A 3385 bp <i>carB</i> gene
carB.R	TTTGAGCTCCGCTTGACCGACTTCAGGTGTTCCA	
<i>wxaco</i> .p.F	TTTCTCGAGGGTTCTACATGGCCGATACG	A 500 bp <i>wxaco</i> gene promoter
<i>wxaco</i> .p.R	CTCAAGCTTGCTGCCTGATTCATCGATGCC	

Appendix D Primers used for real time PCR analysis in this study

Genes	Sequence (5'-3')		Product sizes
	Forward primer	Reverse primer	
<i>hrpG</i>	ATGAACGACCACTCTCCCCCAACG	GAGGCTGGCGTTGACCTGCGAGAC	87 bp
<i>hrpX</i>	GCGGATCGCCAATGCGCTGCGTCTG	GCGGCATCTCCTGGCGCAGCGTGG	114 bp
<i>hpa2</i>	CCCCTACTGCAGCCCAGGTATTTCC	AGCTACGGAATCTCCAGGGACGCAT	115 bp
<i>hpa1</i>	CTTCTTTCAGGTTGACCCAGCCAG	TGCTCGGCATTGTTGCTCTGCTGAA	138 bp
<i>hrcC</i>	ACCGAGCAACGGAATCTTCGACAGG	TGCAGGATACGCTACGATGCCGAC	76 bp
<i>hrcT</i>	AATTGCGCGATGAAGACGCTGGT	CGAACCGTCAATTTGAGTCAGCCA	87 bp
<i>hrpB7</i>	ATCGCGTCTTCTTCTCCGCTGCC	GTGTATGCAGACAAAGCCGCCAGT	77 bp
<i>hrcN</i>	ACGAGCGATCGGAGGTTGCACA	AGTTCATCGAGCTCATTCTTGCGC	75 bp
<i>hrpB5</i>	CAGGGTTCGTGTCCGAGGATGATGTG	GATGAGTGGAACGAATCCGGCCTG	127 bp
<i>hrpB4</i>	TGATCGGCATCGCGCGTAACA	TTGTATCGAGCGCGAGCGTCTGGA	122 bp
<i>hrcJ</i>	CGCGCGCTGAGATACCTGGTGG	CATGTCGTTTCGCATCGTTTTCCGGTG	102 bp
<i>hrpB2</i>	CGCACGCCATCGTTCTGCACATC	GCTTTCAAGCGCTGATGCAGTCCTC	109 bp
<i>hrpB1</i>	GCGAACAGGCAGCAGGCGTACAA	TGGACACGTTTCGATGCATGGATTTC	133 bp
<i>hrcU</i>	CCGATGATCATTGGCCTTGCCTACC	AGCAGCACGCCAATGCGAATA	95 bp
<i>hrcV</i>	TGGTCAACATCCTGGCCGGCAT	AGTGAGGCGATCTGCGACACCATG	123 bp
<i>hpaP</i>	GAGCTGGGAAGCCTGGCTGGATATC	GCGAACTGGTATTGAAGCGAAGCGA	107 bp
<i>hrcQ</i>	TGCTACTGAACGAGGACGACCACGC	CGAGGCGATCGGCCAGCAATAT	69 bp
<i>hrcR</i>	CGCACAACAGATCTGGCCCAAGGA	TCAATTCGCTGAGCGTGAAGGCC	92 bp
<i>hrcS</i>	GTTGCTGCTCTGCCTCAAGGTCTCC	CACCAGCTTGAGCGCGAACGAA	130 bp
<i>hpaA</i>	AACGAAGCCAAGCGCGAGTGCATA	CACTGCCGTGCTGATCGCCCT	100 bp
<i>hrpD5</i>	ACGCAGGTGCATCGTTATGATCCAG	TAACGGACGCTGAGCGCGGGAT	110 bp
<i>hrpD6</i>	GGATCCTTCTGGCGAGCGGCTGC	ACGGCATTGAAGTCGTTGCGTGAGG	120 bp
<i>hrpE</i>	GGTGTGTCCGGTGAATCTCTGGTG	GTTGTTCATGGACTTCTGGGCCTCG	114 bp
<i>hpaB</i>	TCTATCTCCACGAGCCGACGCCA	GGCAGTCCGTAGGAGATGCGCAGAT	82 bp
<i>hrpF</i>	ACCGGATCTGAAGAAGGCATTGACG	CTTTGATCTTGCCGCCGCACTTG	101 bp
<i>hpaF</i>	CACGACTGCCCGCTCAATGCGATAT	TCGCGGATTTGCAGCAATTGGC	70 bp
<i>gyrA</i>	TGATGGCCTCAAGCCTGTGCACCGG	GCCGACGATACGCGCCGACTTGAAG	100 bp